

Bacterial Synthesis of Nano-crystalline Hydroxyapatite

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Abstract

Hydroxyapatite (HA) is a well known candidate for many applications in dentistry and medicine such as bone replacement and regeneration and coatings for medical implants. Nano-crystalline HA exhibits improved mechanical properties and biocompatibility. To optimize the benefits of nano-sized precursors, the particles must be of a uniform shape and size and have minimum degree of agglomeration. The aim of this study was to synthesize of nano-crystalline HA via the biomineralization route. For this purpose, an Iranian strain of *Serratia* (*Serratia marcescens* PTCC 1187) was utilized for the synthesis of nano-crystalline HA. The strain was cultivated. Then the pellet of *S. marcescens* PTCC 1187 was separated and exposed to Glycerol 2-phosphate and Calcium chloride. After 14 days of incubation at 37 °C, the white precipitated material was separated. After drying and calcination at 600 °C the powder was characterized using scanning electron microscopy (SEM), transmission electron microscopy (TEM), X-ray diffraction (XRD) and fourier transform infrared spectroscopy (FTIR) techniques. The results showed that nano-structured HA powder was synthesized and the crystallinity of the powder was relatively high according to the standard. The particles of the powder were single crystal with the size of 25-30 nm. Moreover, the shape and size of the particles were relatively uniform and the agglomeration was lower comparing to the conventional methods. This powder could be used in the regeneration of bone defects, fabrication of medical, dental implants and also as a vector for pharmaceuticals and biological materials such as the genes.

Keywords: Hydroxyapatite; Phosphatase; *Serratia*; Biomineralization; Nanobiotechnology

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°C

(TEM)

(SEM)

nm

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¹ Calcium Phosphate

⁵ Bone Tissue Engineering Scaffolds

⁹ Fracture Toughness

¹³ Vitronectin

¹⁷ Biomineralization

² Hydroxyapatite

⁶ Bioactivity

¹⁰ Wettability

¹⁴ Agglomeration

¹⁸ Strain

³ Biological Fixation

⁷ Nano-crystalline

¹¹ Osteoblast

¹⁵ Sol-gel

¹⁹ Serratia

⁴ Implants

⁸ Sintering

¹² Alkaline Phosphatase

¹⁶ Electrochemical Precipitation

²⁰ Phosphatase

PTCC 1187

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(PTCC

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(OD) nm

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/ ()

°C

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°C

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°C

nm

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() (CaCl₂)

mmol

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mmol ()

mmol

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() (G₂P)

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mmol

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²¹ *Serratia marcescens*

²⁵ Para-nitro Phenyl Phosphate

²⁹ Phosphatase Enzyme Activity Index

³³ TRIS Buffer

³⁷ Centrifuge

²² Persian Type Culture Collection

²⁶ Incubation

³⁰ Calcium Chloride

³⁴ Hydrochloric Acid

³⁸ Filter Paper

²³ Nutrient Broth

²⁷ Optical Density

³¹ Tri-Sodium Citrate

³⁵ Suspension

³⁹ Calcination

²⁴ Pompei

²⁸ Spectrophotometer

³² Glycerol 2-Phosphate

³⁶ Shaking

⁴⁰ Crystallinity

pH

pH

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(XRD)

(TEM)

(SEM)

(FTIR)

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PTCC 1187

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PTCC 1187

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(ASTM)

%

°C

⁴¹ Autoclave

⁴⁴ **T**ransmission **E**lectron **M**icroscopy

⁴⁷ Pang and Bao

⁵⁰ Extracellular Polymeric Matrix

⁵³ Phases

⁵⁶ **A**merican **S**tandard for **T**esting and **M**aterials

⁴² **X**-**R**ay **D**iffraction

⁴⁵ **F**ourier **T**ransform **I**nfrared **S**pectroscopy

⁴⁸ Sata

⁵¹ Peak

⁵⁴ Tri-Calcium Phosphate

⁴³ **S**canning **E**lectron **M**icroscopy

⁴⁶ Williamson-Hall

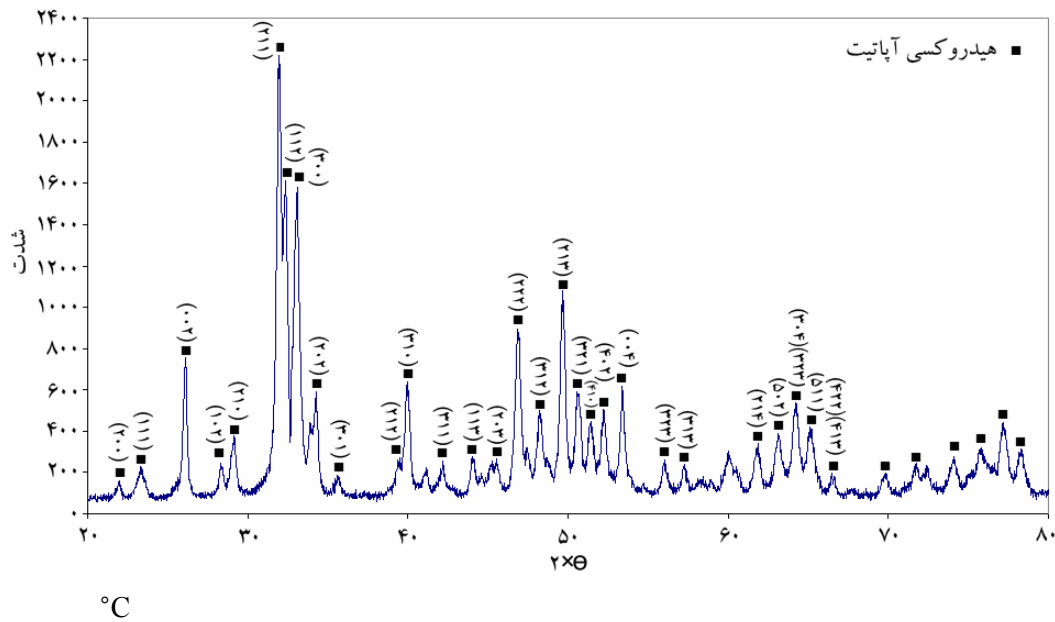
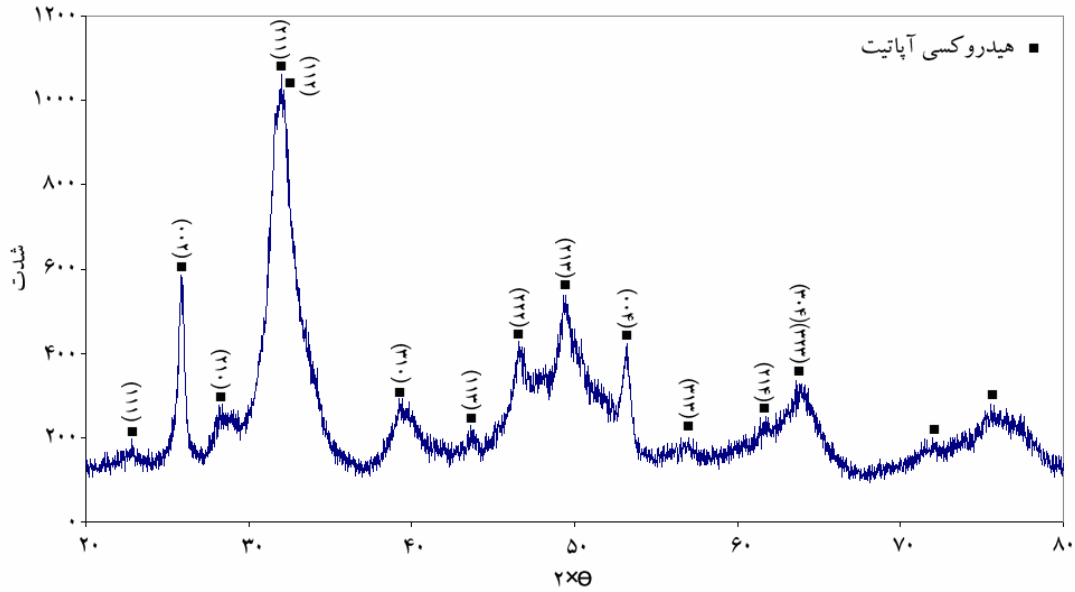
⁴⁹ Enterobacteriaceae

⁵² Thackray

⁵⁵ Calcium Oxide

PTCC

| PTCC | (CFU/ml) | | (ml/CFU) | | (ml/CFU) |
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nm °C

$X_c = -(V / I)$ ()

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cm⁻¹) OH-

cm⁻¹) PO₄³⁻

CO₃²⁻

HPO₄²⁻

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B

CO₃²⁻

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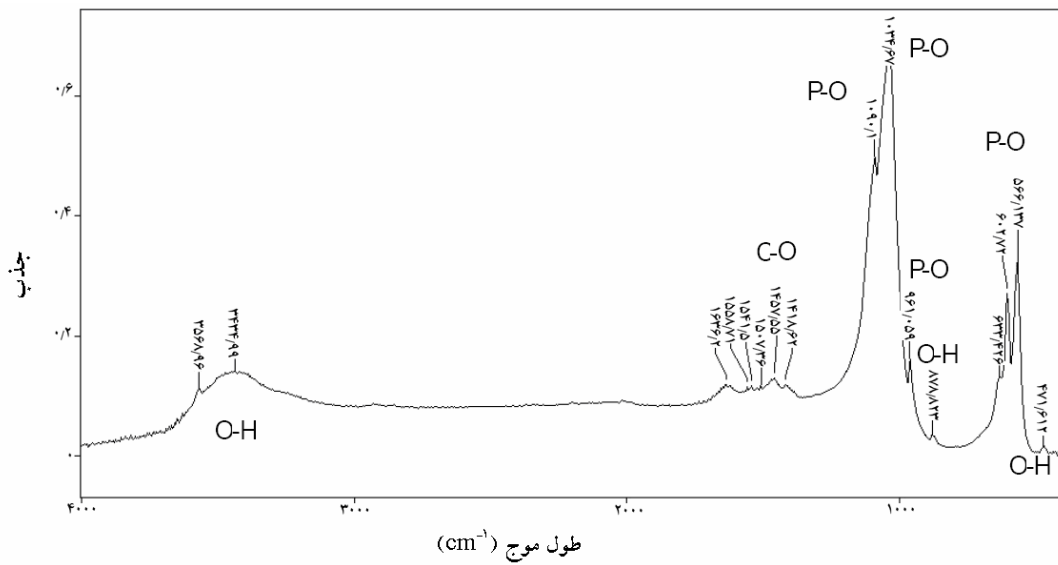
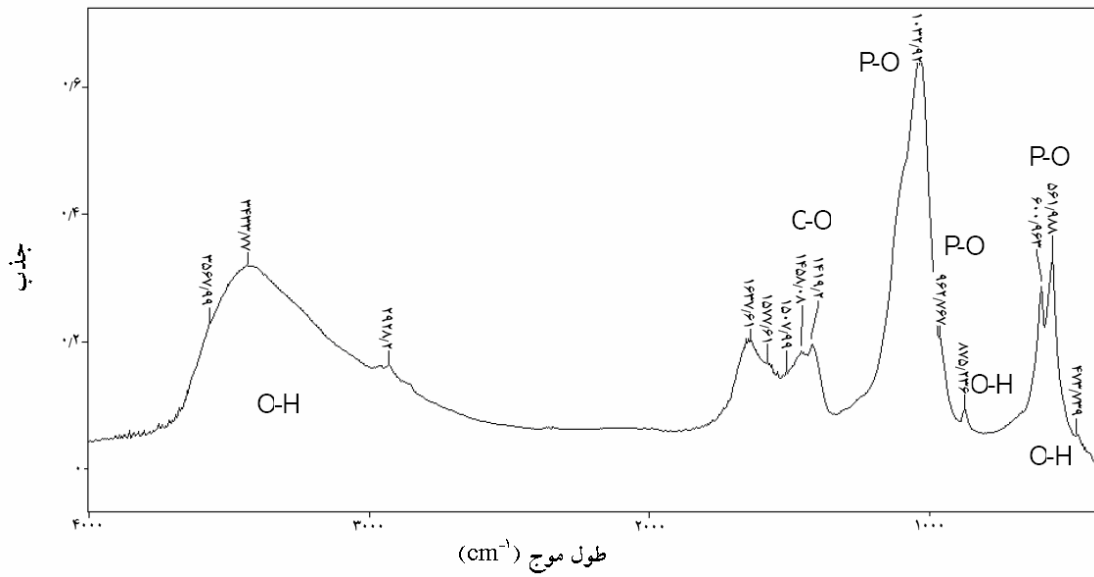
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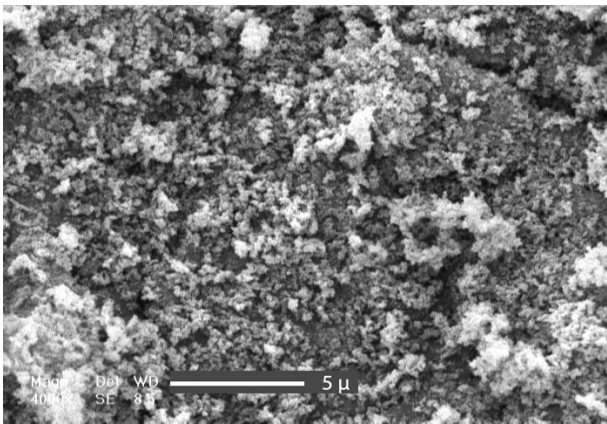
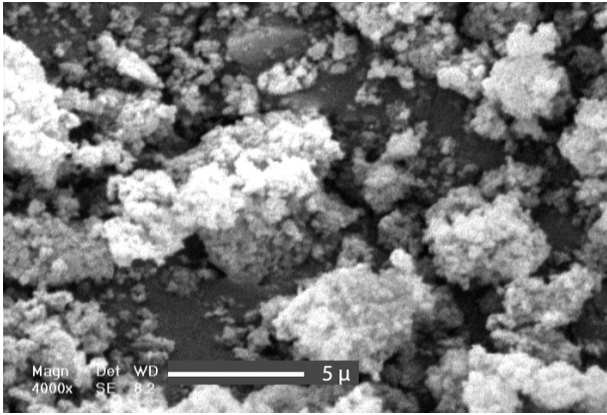
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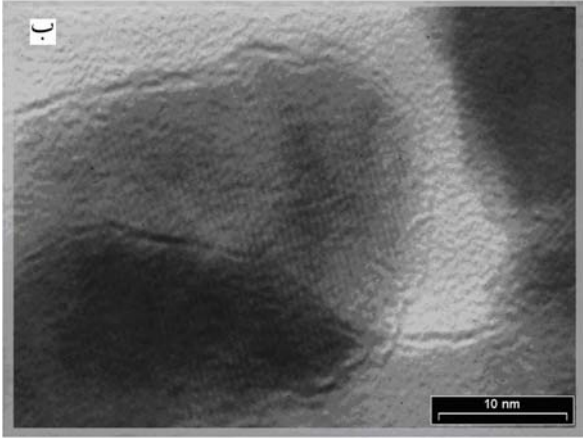
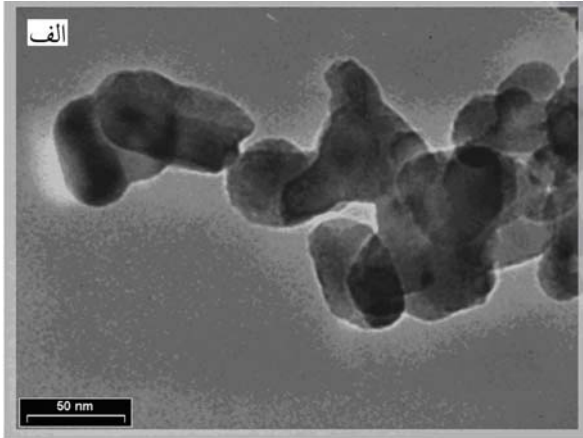
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